


REVIEW

Open Access

# How COVID-19 induces cytokine storm with high mortality



Shintaro Hojyo<sup>1†</sup>, Mona Uchida<sup>1†</sup>, Kumiko Tanaka<sup>1</sup>, Rie Hasebe<sup>1</sup>, Yuki Tanaka<sup>1</sup>, Masaaki Murakami<sup>1\*</sup>   
and Toshio Hirano<sup>1,2\*</sup>

## Abstract

The newly emerging coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was first reported in Wuhan, China, but has rapidly spread all over the world. Some COVID-19 patients encounter a severe symptom of acute respiratory distress syndrome (ARDS) with high mortality. This high severity is dependent on a cytokine storm, most likely induced by the interleukin-6 (IL-6) amplifier, which is hyper-activation machinery that regulates the nuclear factor kappa B (NF- $\kappa$ B) pathway and stimulated by the simultaneous activation of IL-6-signal transducer and activator of transcription 3 (STAT3) and NF- $\kappa$ B signaling in non-immune cells including alveolar epithelial cells and endothelial cells. We hypothesize that IL-6-STAT3 signaling is a promising therapeutic target for the cytokine storm in COVID-19, because IL-6 is a major STAT3 stimulator, particularly during inflammation. We herein review the pathogenic mechanism and potential therapeutic targets of ARDS in COVID-19 patients.

## Background

Coronavirus disease 2019 (COVID-19), which is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has globally spread to an ongoing pandemic since the first case of infection was reported in 2019. Patients with poor prognostic features upon hospital admission frequently encounter complications with significant mortality, particularly by acute respiratory distress syndrome (ARDS) with a broad spectrum of diseases such as multiorgan failure, and blood clots [1]. No effective vaccine strategy or approved medication for the treatment of this contagious disease has been established, although clinical trials are intensively being performed ([https://clinicaltrials.gov/ct2/who\\_table](https://clinicaltrials.gov/ct2/who_table)).

Accumulating evidence suggests that the severity of COVID-19 is associated with an increased level of inflammatory mediators including cytokines and

chemokines such as interleukin (IL)-2, IL-7, IL-10, tumor necrosis factor (TNF), granulocyte colony-stimulating factor (G-CSF), monocyte chemoattractant protein-1 (MCP1; also known as CCL2), macrophage inflammatory protein 1 alpha (MIP1 $\alpha$ ; also known as CCL3), CXC-chemokine ligand 10 (CXCL10), C-reactive protein, ferritin, and D-dimers in blood upon SARS-CoV-2 infection [2–10]. Of note, among the elevated inflammatory mediators, the blood IL-6 level is highly correlated with the disease mortality when COVID-19 survivors and non-survivors are compared [1, 11], suggesting that fatal COVID-19 is characterized as a cytokine release syndrome (CRS) that is induced by a cytokine storm with high mortality [12–14]. Thus, IL-6 serves as a possible mechanism of treatment for severe COVID-19 patients, raising the possibility that one therapeutic option for the disease may be targeting excessive inflammation caused by IL-6 receptor (IL-6R) signaling with monoclonal antibody therapy or treatment with chemical modulators to block the signaling cascade while maintaining a sufficient antiviral primary immune response. In this regard, the use of two clinically approved IL-6R antagonists, tocilizumab (TCZ) and

\* Correspondence: [murakami@igm.hokudai.ac.jp](mailto:murakami@igm.hokudai.ac.jp); [hirano-toshio@qst.go.jp](mailto:hirano-toshio@qst.go.jp)

<sup>†</sup>Shintaro Hojyo and Mona Uchida contributed equally to this work.

<sup>1</sup>Molecular Psychoimmunology, Institute for Genetic Medicine, Graduate School of Medicine, Hokkaido University, Hokkaido 060-0815, Japan  
Full list of author information is available at the end of the article

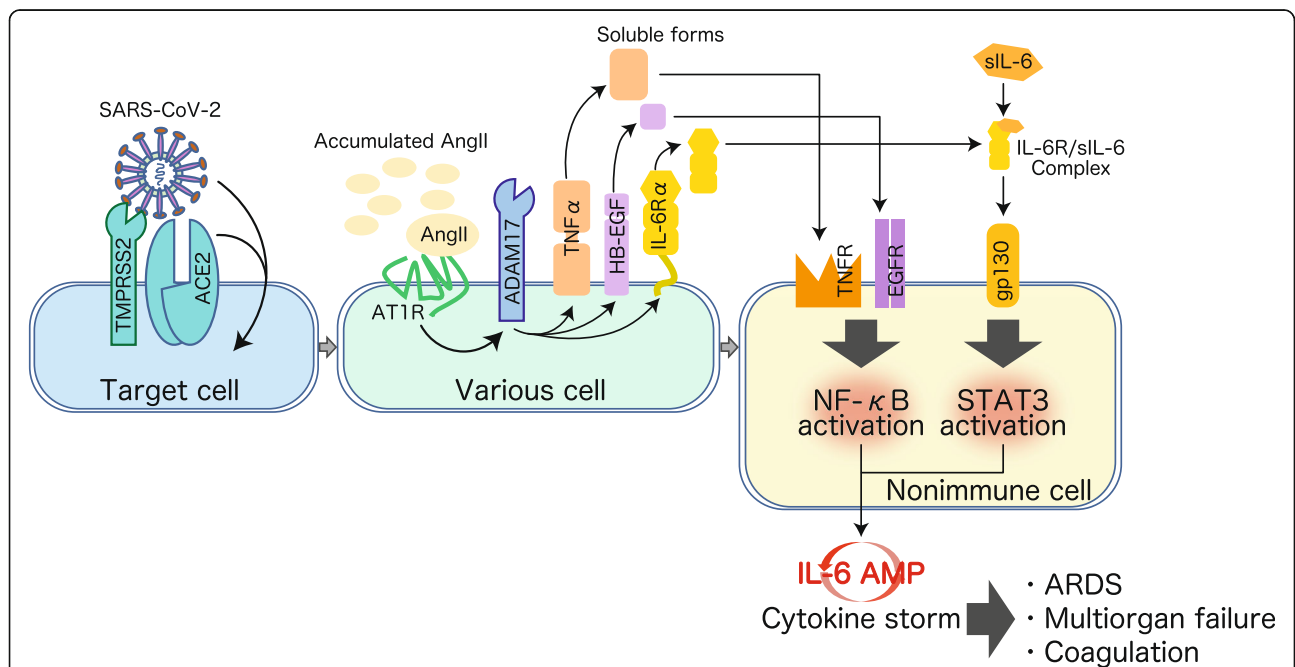


sarilumab (SAR), which are currently used for the treatment of rheumatoid arthritis, could be expected to play a crucial part in the treatment for severely ill patients. With this in mind, here we discuss the potential pathogenetic mechanisms and therapeutic options for COVID-19, focusing on IL-6-signal transducer and activator of transcription 3 (STAT3) signaling.

**Viral entry of SARS-CoV-2**

Within the past two decades, severe respiratory diseases were caused by zoonotic infections of SARS-CoV and MERS-CoV from animals to humans in endemic areas. In late December 2019 in Wuhan City, China, SARS-CoV-2, belonging to the novel RNA *Betacoronavirus*, was discovered as a causative agent of COVID-19, which has currently become a worldwide pandemic. Genomic analysis revealed that SARS-CoV-2 shares about 80% genetic identity with SARS-CoV, around 50% with MERS-CoV, and around 90% with bat-SL-CoVZC45 and bat-SL-CoVZXC21 coronaviruses, suggesting bat-to-human zoonotic transmission of this new virus [15, 16]. Furthermore, proteomic characterization also showed

that SARS-CoV-2 has seven conserved non-structural domains similar to SARS-CoV, suggesting a relationship between the two *Betacoronaviruses*. Despite amino acid differences with SARS-CoV, SARS-CoV-2 has a similar receptor-binding domain to that of SARS-CoV by utilizing angiotensin-converting enzyme II (ACE2) for host cell entry (Fig. 1) [17, 18]. In fact, two independent research groups have provided evidence that SARS-CoV-2 requires ACE2 to infect host cells [16, 19]. Zhou et al. revealed that SARS-CoV-2 can enter cells expressing ACE2 originated from humans, Chinese horseshoe bats, civet, and pigs [16]. Also, it cannot enter cells expressing either dipeptidyl peptidase 4 or aminopeptidase N, the entry receptors for MERS-CoV and HCoV-229E, respectively. SARS-CoV-2 entry via human ACE2 is dependent on transmembrane serine protease 2 (TMPRSS2) and the endosomal cysteine proteases cathepsin B and L (CatB/L) for viral spike (S) protein priming [19]. TMPRSS2 is necessary for cleaving the viral envelope-located trimeric S protein at the S1/S2 and the S2' sites, leading to the fusion of the viral and cellular membranes mediated by the S2 subunit of S protein after the



**Fig. 1** IL-6-STAT3 signaling is a potential therapeutic target for COVID-19 mediated by cytokine storm. SARS-CoV-2 enters cells via cell membrane-localized ACE2 depending on TMPRSS2 and CatB/L activities for viral S protein priming. The viral pathogen-associated molecular patterns trigger IL-6 production followed by activation of the NF-κB pathway via PRRs in both immune and non-immune cells, leading to an inflammatory response. On the other hand, upon the occupancy of ACE2 by SARS-CoV-2, the increased serum level of free Ang II due to a reduction of ACE2-mediated degradation also promotes activation of the NF-κB pathway via AT1R, followed by IL-6 production. Simultaneously, Ang II-AT1R signaling activates ADAM17 and ADAM10 protease activity, and the resulting production of TNF-α, sIL-6Rα, and EGF initiates the TNFR-NF-κB, IL-6R-STAT3, and EGF-NF-κB signaling pathways. Consequently, the concomitant inflammatory cascades of NF-κB- and STAT3-mediated signaling further augment NF-κB activity and establish an inflammatory circuit, the IL-6 amplifier (IL-6 AMP), which describes an IL-6-based positive feedback loop for inflammation in non-immune cells. Thus, the cytokine storm caused by the hyper-activation of NF-κB in IL-6 AMP may cause fatal symptoms such as ARDS, severe pneumonia, multiorgan failure, and coagulation in a subgroup of hospitalized COVID-19 patients. In line with this, the blockade of IL-6-STAT3 signaling should shed light on the treatment of severe COVID-19

engagement of the S1 subunit to the cell surface receptor and for the subsequent viral internalization in the pulmonary epithelium [19]. Notably, ammonium chloride, an inhibitor of CatB/L, inhibited SARS-CoV-2-S protein-driven entry into 293 T cells (TMPRSS2-negative) expressing ACE2, but less so into Caco-2 cells (TMPRSS2-positive). A clinically proven TMPRSS2 inhibitor, camostat mesylate (NCT04321096), which is approved for clinical use in Japan to treat pancreatic inflammation, partially prevented SARS-CoV-2-driven entry into Caco-2 cells, but together with E-64d, an inhibitor of CatB/L completely inhibited the entry. In addition, the forced expression of TMPRSS2 rescued the SARS-CoV-2-S-dependent entry into CatB/L-suppressed 293 T cells, suggesting that the entry of SARS-CoV-2 is induced when cells express TMPRSS2 regardless of CatB/L expression and that SARS-CoV-2 cell entry depends on surface ACE2 and TMPRSS2 molecules. Thus, ACE2 and TMPRSS2 could be predominant targets for COVID-19 therapeutics. Of note, despite the genetic identity of the receptor-binding domain (RBD) structure between SARS-CoV and SARS-CoV-2, several reports have shown anti-RBD monoclonal antibodies cannot neutralize SARS-CoV-2 [20, 21], suggesting that cross-neutralization protection between the two viruses is limited. Moreover, SARS-CoV-2 can infect cells that have Fc receptors (FcRs), which provide the ability of antibody-mediated internalization in macrophages, monocytes, or B cells even without ACE2 and TMPRSS2 expression particularly during the later time point after infection [22].

### **Hyper inflammatory pathogenesis of COVID-19**

Among coronaviruses in humans, HCoV-229E, HCoV-NL63, HCoV-OC43, and HCoV-HKU1 cause mild cold symptoms upon infection in the upper respiratory tract [23]. On the other hand, SARS-CoV, SARS-CoV-2, and MERS-CoV cause fatal pneumonia in some cases after replicating in the lower respiratory tract. Following the infection of SARS-CoV-2 in patients, symptoms occur within 5–6 days when the viral load reaches a peak, and 97.5% of symptomatic patients further develop COVID-19 within 2 weeks [2, 7, 9, 24–26]. Similar to SARS- and MERS-CoV-induced symptoms, a subgroup of patients admitted to hospitals exhibit severe respiratory failure with dyspnea as observed in chest CT scans, bilateral lung infiltration, lymphopenia, muscle/joint pain, headache/dizziness, diarrhea, nausea, and coughing up of blood [3, 4, 6, 8, 10, 27–29]. Notably, severe COVID-19 is associated with mortality and poor prognostic features even when patients are hospitalized [30]. The wide spectrum of the disease severity for these hospitalized patients includes not only pneumonia, severe damage to the airways, and pulmonary edema, but also ARDS, which is a causative syndrome of death in 70% of fatal

COVID-19 cases, in which aggressive inflammatory responses occur [6, 31].

Upon viral infection, alveolar epithelial cells, macrophages, and blood circulating monocytes are activated via toll-like receptors as pattern recognition receptors (PRRs) by the virus products and produce a robust amount of inflammatory cytokines and chemokines, which attract more immune cells, in particular, monocytes and T cells, resulting in widespread lung inflammation (Fig. 1). The postmortem pathology of COVID-19 patients shows interstitial mononuclear inflammatory infiltrates dominated by lymphocytes in the lung and severe lymphopenia with hyperactivated T cells in the peripheral blood [32]. Furthermore, COVID-19 patients also have lower levels of regulatory T cells, which are more obviously decreased in severe cases [33]. Hospitalized patients with severe COVID-19 show high levels of IL-2, IL-7, IL-10, G-CSF, TNF, CXCL10, MCP1, and MIP1 $\alpha$  in serum [6], suggesting that severe COVID-19 is dictated as a cytokine release syndrome (CRS), which is a disorder induced by cytokine storms [12–14]. Among the elevated levels of inflammatory mediators in COVID-19 patients, the blood levels of IL-6 are noticeably higher in non-survivors compared to survivors [1, 2, 11, 34] and predict the need for mechanical ventilation [35].

These findings have led to the hypothesis that the main cause of death of COVID-19 is ARDS with cytokine storms. Notably, intravascular coagulation is one of the causes of multiorgan injury, which is mainly mediated by inflammatory cytokines, in particular, IL-6 [36–38]. Patients exhibit multiorgan failure with coagulation abnormalities represented by lower platelets count and increased D-dimer, which are increasingly associated with poor prognosis and explain the microthrombi of the lungs, lower limbs, hands, brain, heart, liver, and kidneys [39–42]. Similar observations are seen in most patients with SARS-CoV infection who progressed to renal failure [2, 43]. Another reason of multiorgan failure is that SARS-CoV-2 infection in endothelial cells also causes cell death, which leads to vascular leakage and induces a cytopathic effect on airway epithelial cells [44].

Thus, it would seem that the disease severity or mortality comes from cytokine storms including ARDS triggered by viral lung infection, which accounts for multiorgan failure across the body [45]. These inflammatory mediators can also lead to vascular hyperpermeability and stimulate endothelial cells that express ACE2 on arteries and veins that together with viral particles cause systemic inflammation [46, 47].

### **IL-6-STAT3 signaling as a potential cause of the ARDS via cytokine storms in COVID-19 patients**

**IL-6 amplifier, machinery for excessive inflammation**  
SARS-CoV-2 infection induces the endocytosis of ACE2 together with SARS-CoV in target cells including

epithelial cells and endothelial cells, resulting in an increase of serum angiotensin II (Ang II) levels due to the reduction of ACE2 surface expression (Fig. 1) [17, 48]. Ang II increment is also obtained in lung-injury models, in which ACE2 is dramatically reduced upon acid treatment [49]. Ang II acts not only as a vasoconstrictor but also as a pro-inflammatory cytokine via Ang II type 1 receptor (AT1R) [50]. Therefore, it is hypothesized that a renin-angiotensin system (RAS) may be involved in the ARDS development following SARS-CoV-2 infection [12]. In fact, treating mice with AT1R inhibitors or exogenous recombinant ACE2 suppresses ARDS development induced by SARS-CoV infection [17]. In addition, a possible benefit of RAS inhibitors in COVID-19 patients has been reported [51–53]. The Ang II-AT1R signaling axis activates ADAM metallopeptidase domain 17 (ADAM17), which in turn digests the membrane forms of epidermal growth factor family members (EGF, epiregulin, amphiregulin, transforming growth factor- $\alpha$ , etc.) and TNF- $\alpha$ , all of which stimulate the NF- $\kappa$ B pathway (Fig. 1) [12, 50, 54]. ADAM17 is also an enzyme that processes membrane-bound IL-6R $\alpha$  to the soluble form (sIL-6R $\alpha$ ) together with ADAM10. Therefore, we hypothesize serum Ang II and sIL-6R $\alpha$  might be predictive markers of COVID-19 severity.

Once sIL-6R $\alpha$  is generated, the sIL-6R-IL-6 complex transduces intracellular signaling via its binding to gp130, a signal transducer of IL-6, which is expressed on non-immune cells including endothelial cells, epithelial cells, and fibroblasts even without membrane IL-6R expression, followed by the activation of Janus kinase (*JAK*)/*STAT3* [55]. Thus, Ang II-AT1R signaling can create an IL-6-mediated positive feedback loop of NF- $\kappa$ B signaling, a mechanism known as the IL-6 amplifier, during lung inflammation followed by ARDS with multi-organ failure and coagulation (Fig. 1).

The IL-6 amplifier is a hyper NF- $\kappa$ B activation machinery in non-immune cells induced by the simultaneous activation of NF- $\kappa$ B and *STAT3*. It induces a massive and sustained production of NF- $\kappa$ B target genes, including IL-6, chemokines, and growth factors, which is critical for the development of various disease models including lung transplantation, rheumatoid arthritis, and multiple sclerosis [56–60]. Furthermore, we have shown that the co-activation of NF- $\kappa$ B and *STAT3*, which is evidence of activation of the amplifier, is observed in clinical specimens from patients with inflammatory diseases [56, 61]. Additionally, the expression of target molecules of the inflammation amplifier is higher in the serum of patients with rheumatoid arthritis or multiple sclerosis [56, 61]. Moreover, the amplifier activation depends on the concentrations of NF- $\kappa$ B stimulators and of IL-6 around non-immune cells, but these concentrations vary between cells. Indeed, activation tends to

occur more easily in tissue-specific non-immune cells such as tracheal basement cells, synovial fibroblasts, keratinocytes, kidney tubule cells, and chondrocytes. Therefore, through the IL-6 amplifier, these cells could regulate several tissue specific-inflammatory diseases [56–60]. Furthermore, activation of the IL-6 amplifier depends on various environmental and genetic factors. Moreover, we have reported that stress and pain can be triggered for the activation of the IL-6 amplifier at specific blood vessels [62, 63], and some SNPs have an effect on the activation, particularly via the NF- $\kappa$ B pathway [56, 61]. These factors could explain the mild to severe COVID-19 phenotypes observed.

#### Possible therapeutics for COVID-19

As mentioned above, excessive IL-6 levels are highly correlated with the lethal complications of COVID-19 patients [16, 64, 65]. Notably, a previous report showed that inhibition of the NF- $\kappa$ B pathway in animals infected with SARS-CoV decreases mortality and IL-6 levels [66].

As described above, the IL-6 amplifier plays a critical role in chronic inflammatory diseases. The activation of the IL-6 amplifier may induce a cytokine storm, a phenotype of dysregulated inflammation. If this is the case, the cytokine storm in severe COVID-19 could be inhibited by blockade of the IL-6 amplifier [12]. Consistently, chimeric antigen receptor (CAR)-T cell-induced fatal cytokine storm was prevented by an IL-6-*STAT3* blocker [11, 12, 67].

Because the IL-6 amplifier is activated by the coactivation of NF- $\kappa$ B and *STAT3* in non-immune cells, NF- $\kappa$ B and *STAT3* should be potential regulators of the COVID-19-mediated cytokine storm shown in Fig. 1. There are many NF- $\kappa$ B activators, such as PRRs, AT1R, ADAM17/10, TNF- $\alpha$ -TNFR, and EGF-EGFR. Some could be therapeutic targets for the cytokine storm. However, IL-6 is the major *STAT3* activator during inflammatory responses. Considering the availability of IL-6 inhibitors, IL-6-*STAT3* blockade could be a simple option to prevent COVID-19-induced cytokine storm. TCZ is a recombinant humanized monoclonal anti-IL-6R antibody and currently approved for use in patients with CAR-T cell-induced severe cytokine storm or CRS with fever, hypoxia, acute renal failure, hypotension, and cardiac arrhythmia that often warrants ICU admission [11, 67]. The effectiveness of TCZ has been also reported in CRS related to several other conditions, such as sepsis, graft-versus-host disease, and macrophage activation syndrome [68–70]. Common side effects of TCZ include increased upper respiratory tract infections. Therefore, it might be difficult to use TCZ for COVID-19 patients at the early phase of the disease. Nevertheless, the inhibition of the signal transduction mediated by IL-6, the critical *STAT3* inducer for the IL-6 amplifier, through binding to both mIL-6R $\alpha$

and sIL-6R $\alpha$  is a reasonable approach to consider for treating cytokine storm in COVID-19 patients [12, 55]. In fact, Luo and colleagues reported that TCZ administration stabilizes clinical outcomes with a reduction of increased C-reactive protein levels in more than half of COVID-19 patients at risk of a cytokine storm [71]. Moreover, Xu et al. investigated whether TCZ treatment successfully improves the respiratory function in a total of 21-hospitalized COVID-19 patients. Indeed, among these 21 patients, 20 patients were recovered after the TCZ therapy [72]. Toniati et al. also showed that 77% of patients out of 100 rapidly improved clinical and respiratory status after TCZ treatment [73]. These series of study suggest that IL-6-STAT3 signaling could be a promising target for the clinical intervention of COVID-19.

Several ongoing clinical trials for TCZ in COVID-19 patients have been documented (phase II; NCT04317092, NCT04445272, NCT04377659, phase III; NCT04320615, NCT04330638, NCT04345445) (<https://clinicaltrials.gov/ct2/results?cond=COVID-19>). Another anti-IL-6R antibody, SAR, used for rheumatoid arthritis [74], has been tested in a multicenter, double-blind, clinical phase II/III study in patients with severe COVID-19 (NCT04315298) [75]. Although Roche has not reached significant effects of TCZ in phase III study (NCT04320615), more careful study for patient selection is important. We hypothesize that the timing of the treatment should be important for suppressing the cytokine storm induced by SARS-CoV-2 infection. Because patients with severe COVID-19 symptoms have multiorgan failure due to the high expression level of cytokines, some of which regulate the function of the organs directly or indirectly via blood endothelial cells that express ACE2, we hypothesize that IL-6 inhibitors including TCZ could mitigate cytokine storms in COVID-19 patients before multiorgan failure. As described in the review, the study that found IL-6 inhibitors could suppress cytokine storms in patients after CAR-T cell therapy most likely did so by blocking the IL-6 amplifier [67]. Indeed, on 18th September 2020, phase III study met its primary endpoint, showing that patients with COVID-19 associated pneumonia who received TCZ were 44% less likely to progress to mechanical ventilation or death compared to patients who received placebo. Other therapeutic options that potentially inhibit JAK kinases related to IL-6 signaling, such as baricitinib (phase II/III; NCT04340232) [76], are also potential therapeutic candidates.

## Conclusions

The cytokine storm and CRS in fatal COVID-19 are represented by several pathological features such as ARDS, coagulation, and multiorgan dysfunctions. Since blood IL-6 levels are highly correlated with the lethal

complications of COVID-19, we propose that IL-6 plays a pivotal role in the disease augmentation and can be thus a useful biomarker for determining the disease severity. With this regard, TCZ is a good therapeutic option for disrupting the IL-6 amplifier, an IL-6-mediated hyper-inflammatory mechanism that can be triggered by innate immune signaling upon viral infection together with Ang II-AT1R-mediated signaling due to the reduction of Ang II-quercher ACE2. However, in general, monoclonal antibody-based therapeutics impose a substantial cost burden on patients and society. Therefore, future research should be directed to the identification of responsible molecules that regulate the IL-6 amplifier in severe COVID-19, thereby enabling the development of new clinical interventions using small molecules against these targets. Intriguingly, SARS-CoV, MERS-CoV, and SARS-CoV-2 show high genomic diversity and are considered to have originated from bats. Given that these viruses share viral spike proteins and utilize human ACE2 receptor for their entry, it is highly possible that a novel bat coronavirus may have been transmitted to humans to cause emerging infectious diseases. Therefore, clarification of the molecular mechanism for how CRS promotes multiple symptoms in coronavirus-related diseases is needed. Better understanding of this issue will facilitate the development of novel therapies in preparation for future COVID outbreaks.

## Abbreviations

ACE2: Angiotensin-converting enzyme II; ADAM17/10: ADAM metalloproteinase domain 17/10; Ang II: Angiotensin II; ARDS: Acute respiratory distress syndrome; AT1R: Ang II type 1 receptor; CAR: Chimeric antigen receptor; Cat: Cathepsin; COVID-19: Coronavirus disease 2019; CRS: Cytokine release syndrome; CXCL10: CXC-chemokine ligand 10; EGF: Epidermal growth factor; G-CSF: Granulocyte colony-stimulating factor; HCoV: Human coronavirus; ICU: Intensive care unit; IL: Interleukin; MCP1: Monocyte chemoattractant protein-1; MIP1 $\alpha$ : Macrophage inflammatory proteins 1-alpha; MERS-CoV: Middle East respiratory syndrome; PRR: Pattern recognition receptor; RBD: Receptor-binding domain; SAR: Sarilumab; SARS-CoV: Severe acute respiratory syndrome coronavirus; STAT3: Signal transducer and activator of transcription 3; TCZ: Tocilizumab; TMPRSS2: Transmembrane serine protease 2; TNF: Tumor necrosis factor

## Acknowledgements

Not applicable.

## Authors' contributions

All authors contributed to the preparation of the manuscript. The authors read and approved the final manuscript.

## Funding

Not applicable.

## Availability of data and materials

Not applicable.

## Ethics approval and consent to participate

Not applicable.

## Consent for publication

Not applicable.

**Competing interests**

The authors declare that they have no commercial or financial relationships that could be construed as a potential conflict of interest.

**Author details**

<sup>1</sup>Molecular Psychoimmunology, Institute for Genetic Medicine, Graduate School of Medicine, Hokkaido University, Hokkaido 060-0815, Japan.

<sup>2</sup>Headquarters, National Institutes for Quantum and Radiological Science and Technology, Chiba 263-8555, Japan.

Received: 29 July 2020 Accepted: 18 September 2020

Published online: 01 October 2020

**References**

- Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet*. 2020;395(10229):1054–62.
- Tay MZ, Poh CM, Renia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. *Nat Rev Immunol*. 2020;20(6):363–74.
- Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest*. 2020;130(5):2620–9.
- Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*. 2020;395(10223):507–13.
- Coronaviridae Study Group of the International Committee on Taxonomy of V. The species severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2. *Nat Microbiol*. 2020;5(4):536–44.
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395(10223):497–506.
- Kim JY, Ko JH, Kim Y, Kim YJ, Kim JM, Chung YS, et al. Viral load kinetics of SARS-CoV-2 infection in first two patients in Korea. *J Korean Med Sci*. 2020;35(7):e86.
- Liu Y, Yang Y, Zhang C, Huang F, Wang F, Yuan J, et al. Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. *Sci China Life Sci*. 2020;63(3):364–74.
- Pan Y, Zhang D, Yang P, Poon LLM, Wang Q. Viral load of SARS-CoV-2 in clinical samples. *Lancet Infect Dis*. 2020;20(4):411–2.
- Phan LT, Nguyen TV, Luong QC, Nguyen TV, Nguyen HT, Le HQ, et al. Importation and human-to-human transmission of a novel coronavirus in Vietnam. *N Engl J Med*. 2020;382(9):872–4.
- Liu B, Li M, Zhou Z, Guan X, Xiang Y. Can we use interleukin-6 (IL-6) blockade for coronavirus disease 2019 (COVID-19)-induced cytokine release syndrome (CRS)? *J Autoimmun*. 2020;111:102452.
- Hirano T, Murakami M. COVID-19: a new virus, but a familiar receptor and cytokine release syndrome. *Immunity*. 2020;52(5):731–3.
- Mahmudpour M, Roozbeh J, Keshavarz M, Farrokhi S, Nabipour I. COVID-19 cytokine storm: the anger of inflammation. *Cytokine*. 2020;133:155151.
- McGonagle D, Sharif K, O'Regan A, Bridgewood C. The role of cytokines including interleukin-6 in COVID-19 induced pneumonia and macrophage activation syndrome-like disease. *Autoimmun Rev*. 2020;19(6):102537.
- Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *Lancet*. 2020;395(10224):565–74.
- Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270–3.
- Kuba K, Imai Y, Rao S, Gao H, Guo F, Guan B, et al. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. *Nat Med*. 2005;11(8):875–9.
- Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature*. 2003;426(6965):450–4.
- Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell*. 2020;181(2):271–80 e8.
- Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, Veesler D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell*. 2020;181(2):281–92 e6.
- Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, et al. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *bioRxiv*. 2020.
- Iwasaki A, Yang Y. The potential danger of suboptimal antibody responses in COVID-19. *Nat Rev Immunol*. 2020;20(6):339–41.
- Fehr AR, Perlman S. Coronaviruses: an overview of their replication and pathogenesis. *Methods Mol Biol*. 2015;1282:1–23.
- Lauer SA, Grantz KH, Bi Q, Jones FK, Zheng Q, Meredith HR, et al. The incubation period of coronavirus disease 2019 (COVID-19) from publicly reported confirmed cases: estimation and application. *Ann Intern Med*. 2020;172(9):577–82.
- Peiris JS, Chu CM, Cheng VC, Chan KS, Hung IF, Poon LL, et al. Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. *Lancet*. 2003;361(9371):1767–72.
- Zou L, Ruan F, Huang M, Liang L, Huang H, Hong Z, et al. SARS-CoV-2 viral load in upper respiratory specimens of infected patients. *N Engl J Med*. 2020;382(12):1177–9.
- Chan JF, Yuan S, Kok KH, To KK, Chu H, Yang J, et al. A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster. *Lancet*. 2020;395(10223):514–23.
- de Wit E, van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: recent insights into emerging coronaviruses. *Nat Rev Microbiol*. 2016;14(8):523–34.
- Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, et al. Clinical characteristics of coronavirus disease 2019 in China. *N Engl J Med*. 2020;382(18):1708–20.
- Jose RJ, Manuel A. COVID-19 cytokine storm: the interplay between inflammation and coagulation. *Lancet Respir Med*. 2020;8(6):e46–e7.
- Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan. *JAMA*. China; 2020.
- Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med*. 2020;8(4):420–2.
- Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with COVID-19 in Wuhan. *Clin Infect Dis*. China; 2020.
- Zhang Y, Li J, Zhan Y, Wu L, Yu X, Zhang W, et al. Analysis of serum cytokines in patients with severe acute respiratory syndrome. *Infect Immun*. 2004;72(8):4410–5.
- Herold T, Jurinovic V, Arnreich C, Lipworth BJ, Hellmuth JC, von Bergwelt-Baildon M, et al. Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19. *J Allergy Clin Immunol*. 2020;146(1):128–36 e4.
- Merad M, Martin JC. Pathological inflammation in patients with COVID-19: a key role for monocytes and macrophages. *Nat Rev Immunol*. 2020;20(6):355–62.
- Levi M, Nieuwdorp M, van der Poll T, Strokes E. Metabolic modulation of inflammation-induced activation of coagulation. *Semin Thromb Hemost*. 2008;34(1):26–32.
- van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG. The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol*. 2017;17(7):407–20.
- Liu PP, Blet A, Smyth D, Li H. The science underlying COVID-19: implications for the cardiovascular system. *Circulation*. 2020;142(1):68–78.
- Tang N, Bai H, Chen X, Gong J, Li D, Sun Z. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost*. 2020;18(5):1094–9.
- Xiang-Hua Y, Le-Min W, Ai-Bin L, Zhu G, Riquan L, Xu-You Z, et al. Severe acute respiratory syndrome and venous thromboembolism in multiple organs. *Am J Respir Crit Care Med*. 2010;182(3):436–7.
- Zhang Y, Xiao M, Zhang S, Xia P, Cao W, Jiang W, et al. Coagulopathy and antiphospholipid antibodies in patients with COVID-19. *N Engl J Med*. 2020;382(17):e38.
- Chu KH, Tsang WK, Tang CS, Lam MF, Lai FM, To KF, et al. Acute renal impairment in coronavirus-associated severe acute respiratory syndrome. *Kidney Int*. 2005;67(2):698–705.
- Park WB, Kwon NJ, Choi SJ, Kang CK, Choe PG, Kim JY, et al. Virus isolation from the first patient with SARS-CoV-2 in Korea. *J Korean Med Sci*. 2020;35(7):e84.
- Zhang B, Zhou X, Qiu Y, Song Y, Feng F, Feng J, et al. Clinical characteristics of 82 cases of death from COVID-19. *PLoS One*. 2020;15(7):e0235458.

46. Chen L, Li X, Chen M, Feng Y, Xiong C. The ACE2 expression in human heart indicates new potential mechanism of heart injury among patients infected with SARS-CoV-2. *Cardiovasc Res.* 2020;116(6):1097–100.
47. Hamming I, Timens W, Bulthuis ML, Lely AT, Navis G, van Goor H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J Pathol.* 2004;203(2):631–7.
48. Verdecchia P, Cavallini C, Spanevello A, Angeli F. The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *Eur J Intern Med.* 2020;76:14–20.
49. Imai Y, Kuba K, Rao S, Huan Y, Guo F, Guan B, et al. Angiotensin-converting enzyme 2 protects from severe acute lung failure. *Nature.* 2005;436(7047):112–6.
50. Eguchi S, Kawai T, Scalia R, Rizzo V. Understanding angiotensin II type 1 receptor signaling in vascular pathophysiology. *Hypertension.* 2018;71(5):804–10.
51. Meng J, Xiao G, Zhang J, He X, Ou M, Bi J, et al. Renin-angiotensin system inhibitors improve the clinical outcomes of COVID-19 patients with hypertension. *Emerg Microbes Infect.* 2020;9(1):757–60.
52. Zhang P, Zhu L, Cai J, Lei F, Qin JJ, Xie J, et al. Association of inpatient use of angiotensin-converting enzyme inhibitors and angiotensin II receptor blockers with mortality among patients with hypertension hospitalized with COVID-19. *Circ Res.* 2020;126(12):1671–81.
53. Matsuzawa Y, Ogawa H, Kimura K, Konishi M, Kirigaya J, Fukui K, et al. Renin-angiotensin system inhibitors and severity of coronavirus disease-2019 in Kanagawa, Japan: a retrospective cohort study. *Hypertens Res.* In press.
54. Scheller J, Chalaris A, Garbers C, Rose-John S. ADAM17: a molecular switch to control inflammation and tissue regeneration. *Trends Immunol.* 2011;32(8):380–7.
55. Murakami M, Kamimura D, Hirano T. Pleiotropy and specificity: insights from the interleukin 6 family of cytokines. *Immunity.* 2019;50(4):812–31.
56. Fujita M, Yamamoto Y, Jiang JJ, Atsumi T, Tanaka Y, Ohki T, et al. NEDD4 is involved in inflammation development during keloid formation. *J Invest Dermatol.* 2019;139(2):333–41.
57. Harada M, Kamimura D, Arima Y, Kohsaka H, Nakatsuji Y, Nishida M, et al. Temporal expression of growth factors triggered by epiregulin regulates inflammation development. *J Immunol.* 2015;194(3):1039–46.
58. Lee J, Nakagiri T, Kamimura D, Harada M, Oto T, Susaki Y, et al. IL-6 amplifier activation in epithelial regions of bronchi after allogeneic lung transplantation. *Int Immunol.* 2013;25(5):319–32.
59. Ogura H, Murakami M, Okuyama Y, Tsuruoka M, Kitabayashi C, Kanamoto M, et al. Interleukin-17 promotes autoimmunity by triggering a positive-feedback loop via interleukin-6 induction. *Immunity.* 2008;29(4):628–36.
60. Ota M, Tanaka Y, Nakagawa I, Jiang JJ, Arima Y, Kamimura D, et al. Role of chondrocytes in the development of rheumatoid arthritis via transmembrane protein 147-mediated NF-kappaB activation. *Arthritis Rheumatol.* 2020;72(6):931–42.
61. Murakami M, Harada M, Kamimura D, Ogura H, Okuyama Y, Kumai N, et al. Disease-association analysis of an inflammation-related feedback loop. *Cell Rep.* 2013;3(3):946–59.
62. Arima Y, Kamimura D, Atsumi T, Harada M, Kawamoto T, Nishikawa N, et al. A pain-mediated neural signal induces relapse in murine autoimmune encephalomyelitis, a multiple sclerosis model. *Elife.* 2015;4.
63. Arima Y, Ohki T, Nishikawa N, Higuchi K, Ota M, Tanaka Y, et al. Brain micro-inflammation at specific vessels dysregulates organ-homeostasis via the activation of a new neural circuit. *Elife.* 2017;6.
64. Magro G. COVID-19: review on latest available drugs and therapies against SARS-CoV-2. Coagulation and inflammation cross-talking. *Virus Res.* 2020;286:198070.
65. Giamarellos-Bourboulis EJ, Netea MG, Rovina N, Akinosoglou K, Antoniadou A, Antonakos N, et al. Complex immune dysregulation in COVID-19 patients with severe respiratory failure. *Cell Host Microbe.* 2020;27(6):992–1000 e3.
66. DeDiego ML, Nieto-Torres JL, Regla-Nava JA, Jimenez-Guardeno JM, Fernandez-Delgado R, Fetz C, et al. Inhibition of NF-kappaB-mediated inflammation in severe acute respiratory syndrome coronavirus-infected mice increases survival. *J Virol.* 2014;88(2):913–24.
67. Neelapu SS, Tummala S, Kebriaei P, Wierda W, Gutierrez C, Locke FL, et al. Chimeric antigen receptor T-cell therapy - assessment and management of toxicities. *Nat Rev Clin Oncol.* 2018;15(1):47–62.
68. De Benedetti F, Brunner HI, Ruperto N, Kenwright A, Wright S, Calvo I, et al. Randomized trial of tocilizumab in systemic juvenile idiopathic arthritis. *N Engl J Med.* 2012;367(25):2385–95.
69. Kennedy GA, Varelias A, Vuckovic S, Le Texier L, Gartlan KH, Zhang P, et al. Addition of interleukin-6 inhibition with tocilizumab to standard graft-versus-host disease prophylaxis after allogeneic stem-cell transplantation: a phase 1/2 trial. *Lancet Oncol.* 2014;15(13):1451–9.
70. Sheng F, Han M, Huang Z, Zhang L. Interleukin 6 receptor inhibitor tocilizumab suppresses cytokine expression, inflammasome activation and phagocytosis in a cell model of sepsis. *Pharmazie.* 2016;71(11):636–9.
71. Luo P, Liu Y, Qiu L, Liu X, Liu D, Li J. Tocilizumab treatment in COVID-19: a single center experience. *J Med Virol.* 2020;92(7):814–8.
72. Xu X, Han M, Li T, Sun W, Wang D, Fu B, et al. Effective treatment of severe COVID-19 patients with tocilizumab. *Proc Natl Acad Sci U S A.* 2020;117(20):10970–5.
73. Toniati P, Piva S, Cattalini M, Garrafa E, Regola F, Castellini F, et al. Tocilizumab for the treatment of severe COVID-19 pneumonia with hyperinflammatory syndrome and acute respiratory failure: a single center study of 100 patients in Brescia, Italy. *Autoimmun Rev.* 2020;19(7):102568.
74. Lamb YN, Deeks ED. Sarilumab: a review in moderate to severe rheumatoid arthritis. *Drugs.* 2018;78(9):929–40.
75. Sanders JM, Monogue ML, Jodlowski TZ, Cutrell JB. Pharmacologic treatments for coronavirus disease 2019 (COVID-19): a review. *JAMA.* 2020.
76. Mogul A, Corsi K, McAuliffe L. Baricitinib: the second FDA-approved JAK inhibitor for the treatment of rheumatoid arthritis. *Ann Pharmacother.* 2019;53(9):947–53.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

